

Diverse parentage relationships in paternal mouthbrooding fishes

Abecia, Janine E.; King, Alison J.; Luiz, Osmar J.; Crook, David A.; Wedd, Dion; Banks, Sam C.

Published in:
Biology Letters

DOI:
[10.1098/rsbl.2021.0576](https://doi.org/10.1098/rsbl.2021.0576)

Published: 01/05/2022

Document Version
Publisher's PDF, also known as Version of record

[Link to publication](#)

Citation for published version (APA):

Abecia, J. E., King, A. J., Luiz, O. J., Crook, D. A., Wedd, D., & Banks, S. C. (2022). Diverse parentage relationships in paternal mouthbrooding fishes. *Biology Letters*, 18(5), 1-5. Article 20210576. <https://doi.org/10.1098/rsbl.2021.0576>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Research



Cite this article: Abecia JE, King AJ, Luiz OJ, Crook DA, Wedd D, Banks SC. 2022 Diverse parentage relationships in paternal mouthbrooding fishes. *Biol. Lett.* **18**: 20210576.
<https://doi.org/10.1098/rsbl.2021.0576>

Received: 2 November 2021

Accepted: 14 April 2022

Subject Areas:

ecology, behaviour

Keywords:

paternal, mouthbrooding, genetic parentage analysis, certainty of paternity

Author for correspondence:

Janine E. Abecia

e-mail: janineerica.abecia@cdu.edu.au

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.5958835>.

Diverse parentage relationships in paternal mouthbrooding fishes

Janine E. Abecia¹, Alison J. King^{1,2}, Osmar J. Luiz¹, David A. Crook^{1,2}, Dion Wedd¹ and Sam C. Banks¹

¹Research Institute for the Environment and Livelihoods, Charles Darwin University, Ellengowan Drive, Casuarina, NT 0810, Australia

²Centre for Freshwater Ecosystems, La Trobe University, Albury/Wodonga Campus, Vic 3690, Australia

JEA, 0000-0003-4271-8830

While mouthbrooding is not an uncommon parental care strategy in fishes, paternal mouthbrooding only occurs in eight fish families and is little studied. The high cost of paternal mouthbrooding to the male implies a low risk of investment in another male's offspring but genetic parentage patterns are poorly known for paternal mouthbrooders. Here, we used single-nucleotide polymorphism genetic data to investigate parentage relationships of broods of two mouthbrooders of northern Australian rivers, mouth almighty *Glossamia aprion* and blue catfish *Neoarius graeffei*. For *N. graeffei*, we found that the parentage pattern was largely monogamous with the brooder male as the sire. For *G. aprion*, the parentage pattern was more heterogeneous including observations of monogamous broods with the brooder male as the sire (73%), polygyny (13%), cuckoldry (6%) and a brood genetically unrelated to the brooder male (6%). Findings demonstrate the potential for complex interrelationships of male care, paternity confidence and mating behaviour in mouthbrooding fishes.

1. Introduction

Paternal mouthbrooding is a parental care strategy where the male parent incubates eggs/larvae in the buccal cavity to protect the offspring from biotic and abiotic stressors [1,2]. Mouthbrooding involves a considerable investment from the adult, as the brooder is typically unable to feed, or at least is severely constrained [2,3]. Due to the high level of male investment [4], paternal mouthbrooding is generally assumed to be associated with high confidence in paternity (monoandry) [5]. The only two genetic parentage studies on paternal mouthbrooders, *Osteoglossum bicirrhosum* and *Sphaeramia nematoptera*, reported monogamous broods with the brooder male as the sire (73–92%), but revealed occurrences of multiple paternity (7.6–21%) for both species, multiple maternity (10.6%) for *S. nematoptera* and broods sired by a different male (18%) for *O. bicirrhosum* [6,7]. Investigation of genetic mating patterns is critical for illuminating individual reproductive strategies and behaviours, which may impact the species' reproductive success [8,9]. Because empirical data on the genetic parentage patterns of broods of paternal mouthbrooders are still poorly known, our understanding of the evolutionary drivers of this parental care strategy remains limited.

Male care, the predominant parental care type in fish, is theorized to be correlated with external fertilization, pair-spawning and paternity confidence [10–12]. Fish mating systems are also associated with the type of parental care exhibited [5]. Male care has been shown to correlate with high incidences of multiple maternity but low rates of multiple paternity, while in female and

biparental care, this relationship is generally reversed [5]. Mixed parentages in broods can be a consequence of alternative reproductive behaviours within species such as cuckoldry in the male nesting *Lepomis punctatus* and fry adoption in the maternal mouthbrooding *Simochromis pleurospilus* [8,9,13–15]. For paternal mouthbrooders, broods may be sired by more than one male, by a different male or may consist of eggs from more than one female if some of the eggs received from the first female are partially cannibalized [6,7]. Because all the offspring and the putative father can be genetically sampled [5,6,9], genetic studies on paternal mouthbrooding can test hypotheses on male parental care including sexual selection, mating patterns and paternity confidence.

We investigate patterns of paternity in broods of two freshwater paternal mouthbrooders from divergent fish families, mouth almighty *Glossamia aprion* (Apogonidae) and blue catfish *Neoarius graeffei*. Although these species are fairly poorly studied, *G. aprion* spawns all year round, incubates for two to three weeks and has a fecundity of 104–532 and brood size of 4–416 eggs [16–21]. Contrastingly, *N. graeffei* spawns during the wet season only, incubates for four to five weeks and has a fecundity of 4–128 and brood size of 1–88 eggs [17,18,21]. For both species, mouthbrooding comes at the expense of the parent's feeding, body condition and digestive and respiratory capacities [17]. While *G. aprion* has been observed to form mating pairs in aquaria [22], the detailed reproductive activity (e.g. fertilization step) of both species has never been observed [18]. Based on previous studies [5–7], we predict that the broods of both species are genetically related to the brooder male; support for this prediction will provide justification for paternity confidence in these two study species and for their paternal mouthbrooding strategy. We also predict that the broods of paternal mouthbrooders will show higher rates of multiple maternity relative to multiple paternity.

2. Materials and methods

(a) Sample collection and preparation

Samples of 115 *N. graeffei* (nine brooding males, 90 offspring and 16 non-brooding adults) and 261 *G. aprion* (18 brooding males, 180 offspring and 63 non-brooding adults) were collected across rivers of Northern Territory, Australia. *G. aprion* was collected in five sites: the Mary River (AHMR), Adelaide River (DRAR) and Daly River (OODR, SHFR, GJKR) and *N. graeffei* was collected from one site (Mary River) using electrofishing. Non-brooding males and females were included to provide the basic population genetics context for our study including allele frequency estimates required for parentage analysis. Each fish was euthanized in AQUI-S (175 mg l⁻¹ for 20 min) upon capture; brooding males together with their offspring were bagged individually, labelled and kept in an ice slurry until processed (see electronic supplementary material, S1–S3 for fish collection and justification of method). Fish collections were made as part of a larger study investigating trait variation in freshwater fishes, thereby using specimens for multiple research purposes. These species are abundant, widely distributed in the northern Australian rivers and not considered threatened species [23].

We randomly sampled 9–11 offspring from each brooding male parent. This sample size gave us a 95% chance of detecting the contribution of more than one parent (of either sex) to a brood as long as the dominant parent contributed at least 22–30% of the offspring ([5,8]; see electronic supplementary

material, S4 for explanation). Approximately 5–7 mg of adult muscle, larval tissue or whole egg was collected and stored in 70% ethanol until DNA extraction and genomic sequencing.

(b) Single-nucleotide polymorphism discovery and filtering

Samples were genotyped at single-nucleotide polymorphism (SNP) loci using the DArTSeq method at Diversity Arrays Technology Pty Ltd, Canberra, Australia [24]. We used strict SNP filtering criteria to retain only high confidence genotypes for parentage analysis using a combination of custom R scripts, 'adegenet', 'dartR' and 'plotly' packages in R [25–28] (see electronic supplementary material, S5 for detailed SNPs filtering and genetic diversity estimation methods).

(c) Parentage and sib-ship inference analysis

We first examined the percentage of opposite homozygotes (%OH) between each offspring in the brood and their brooding parent to help identify typing error rates and possible non-paternity of brooding male using the CalcOHLLR function in the 'sequoia' R package [27,29]. We also examined the %OH between non-parents and each offspring in a brood by randomly calculating the OH of each offspring with another brooding male. To determine whether the brooding male parent is the true father of each brood and whether each brood consisted of full- or half-siblings, we performed a full-likelihood parentage analysis using the Colony(v. 2) software [30]. Colony was run using moderate prior, medium run, assumed polygamy for both males and females, and using a range of error rates (0.01–1%) informed by %OH calculations.

3. Results

(a) Single-nucleotide polymorphism discovery and filtering

From 23 963 SNPs, 1748 SNPs (nine brooding males, 90 offspring and 13 non-brooding adults) remained for analysis in *N. graeffei* after filtering with a minimum and mean sequencing depth of 20 and 30.98 per genotype, respectively. From 12 351 SNPs, 1313 SNPs (15 brooding males, 146 offspring and 58 non-brooding adults) remained for analysis in *G. aprion* after filtering with a minimum and mean sequencing depth of 20 and 64.62 per genotype, respectively. See table 1 for SNPs metrics of *N. graeffei* and *G. aprion*. For both species, SNP PIC scores were greater than 0.20 and had 1.19% (*N. graeffei*) and 1.26% (*G. aprion*) missing data. (See electronic supplementary material, S6–S9 for genetic diversity indices of the sampled population.)

(b) Percentage of opposite homozygotes, genetic parentage and sib-ship inference

For *N. graeffei*, the offspring in each brood were assigned as full siblings with the brooder male as the sire. The %OH between offspring-brooding parent pair ranged from 0.81 to 1.98%, where we assume that the brooding male is the parent. This range gives an estimate of the genotyping error rate between the brooding parent and its offspring, assuming that the lower bound is closer to the true error rate and the upper bound due to non-paternity of some offspring. By contrast, the %OH of known non-parents and each offspring range from 6.1 to 10%, which represents the estimated

Table 1. SNPs metrics (average \pm s.d.) of *Neoarius graeffei* and *Glossamia aprion* before and after SNP filtering.

	<i>n</i> loci	call rate	mean sequencing depth	mean allele depth ratio	repeatability
<i>Neoarius graeffei</i>					
SNPs pre-filtering \pm s.d. (all samples)	23963	0.82 \pm 0.22	16.99 \pm 10.05	1.51 \pm 0.52	0.99 \pm 0.02
SNPs post-filtering \pm s.d. (all samples)	1748	0.99 \pm 0.02	30.98 \pm 8.24	1.46 \pm 0.27	1
SNPs post-filtering \pm s.d. (adults only)	1748	0.99 \pm 0.02	30.98 \pm 8.24	1.46 \pm 0.27	1
<i>Glossamia aprion</i>					
SNPs pre-filtering \pm s.d. (all samples)	12 351	0.85 \pm 0.18	37.78 \pm 41.6	1.52 \pm 0.6	0.98 \pm 0.02
SNPs post-filtering \pm s.d. (all samples)	1313	0.98 \pm 0.02	64.62 \pm 40.31	1.30 \pm 0.24	1
SNPs post-filtering \pm s.d. (adults only)	1313	0.98 \pm 0.02	64.63 \pm 40.31	1.29 \pm 0.24	1

%OH of non-paternity. At 1% marker error rate, parentage assignment showed that all broods ($n=9/9$) were assigned to their brooding male parent. All the broods were also assigned as full siblings with an inclusion and exclusion probability range of 0.999–1.00 (figure 1a; electronic supplementary material, S10 and S11). At 0.01% error rate, *N. graeffei* broods were assigned as full siblings but the brooding male was excluded as the sire. We assumed that this is due to genotyping error rate given that the %OH is much lower than that for known non-parents.

For *G. aprion*, the parentage patterns were more heterogeneous between the investigated broods. The %OH between each offspring and its respective brooding parent examined ranged from 0 to 4.3%. Between known non-parents and each offspring, the %OH ranges from 0.15% to 37.75%. Parentage assignment showed that the *G. aprion* broods exhibited heterogeneous mating patterns with an inclusion and exclusion probability range of 0.081–1.00 (figure 1b–e; electronic supplementary material, S10 and S11). At 1% and 0.01% error rates, 11 of the 15 broods (73%) were assigned as full siblings with the brooder male as the sire (figure 1b). Two broods (13%) had two females contributing to the broods' genotype (figure 1c), one brood (6%, GA355) was genetically unrelated to the brooder male (figure 1d), and another brood (6%) was fathered by more than one male (figure 1e).

4. Discussion

While the certainty of paternity is theorized to co-evolve with male care [10–12], there are many reports of male care in fishes with incidences of multiple paternity and alternative reproductive behaviours [5,8,9,13–15]. Unlike other types of male care in fishes, there is still little genetic information available on broods of paternal mouthbrooders [5–7]. In this study, we revealed a largely monogamous genetic mating pattern for the two study species. *N. graeffei* showed monogamous broods with the brooder male as the sire, supporting our prediction that the mouthbrooding male was the true father of the offspring. *G. aprion* broods, however, revealed polygyny, adoption and cuckoldry in a background of a mostly monogamous mating pattern. These results suggest intraspecific variability in the genetic mating pattern of *G. aprion*.

Parental care in mouthbrooders affords a very high cost to the care-giving male [17,31,32], so the overall benefit of care depends on balancing fitness cost with a high certainty in

parentage [11,33]. The broods of both species were largely monogamous which is likely associated with the relative costs and benefits of their paternal mouthbrooding strategy. There are evolutionary incentives for male carers to provide care if they are the only male progenitor of their brood because the fitness returns of caring trades-off against the energy invested [34–36]. The energetic costs associated with paternal mouthbrooding may limit the brooder's ability to mate with more than one female [4,17,31,32]. Moreover, the limited capacity of the buccal cavity to accommodate eggs from another female may also restrict male parents from engaging in extra-pair mating [37,38].

The contrasting reproductive ecologies and level of paternal investment may have contributed to the variations in the parentage patterns observed in the two study species. *N. graeffei* broods fewer eggs (1–88) and for a longer duration (four to five weeks) than *G. aprion* which broods 4–512 eggs for two to three weeks [17,18]. Spawning for *N. graeffei* occurs in the highly interconnected and productive wet season, while *G. aprion* spawns throughout the year, with higher occurrence during the wet–dry seasons, where young are exposed to habitat restriction and concentration of organisms (predators, competition) during the extended dry season [21]. In this study, *G. aprion* had considerable variation in genetic mating behaviour. Variation in mating behaviour within species is suggested to increase the success of passing on multiple gene types in response to varying ecological factors in the environment [39], which seems a plausible justification for *G. aprion*.

The two *G. aprion* broods, (i) fathered by more than one male and (ii) genetically unrelated to the brooder male, could possibly be explained by cuckoldry and adoption of brood, respectively [6–9]. From observations in aquaria, the male and female pair off during courtship; the female then releases the egg mass, and the male begins taking it in its buccal cavity when it is half-way out of the female [22]. A female was also seen to chase the brooding male making it spit its eggs—which the male did not take back [22]. Such behavioural accounts in *G. aprion* likely provide an opportunity for other non-brooding males to take another's brood and care for them, and also for the male to 'swap' broods. While there is apparently little genetic benefit for males to care for the offspring of another, females may preferentially choose males who are already caring for progeny [40].

In broods with multiple maternity, *G. aprion* males could have mated with a female with only a few eggs or cannibalized a portion of their brood when presented with a new mating opportunity. This is likely since the fecundity of

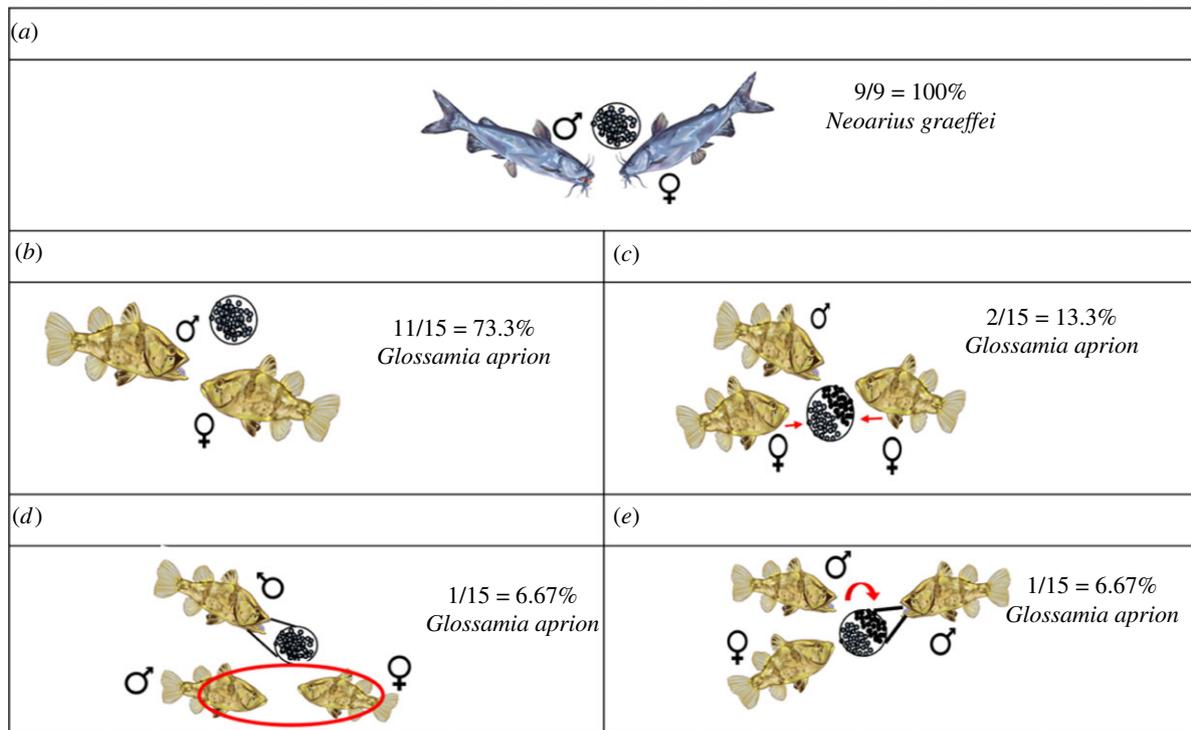


Figure 1. Parentage patterns of *Neoarius graeffei* and *Glossamia aprion* showing (a,b) monogamous brood with the brooder male as the sire: (a) one mother and one father in a brood and (b) one mother and one father in a brood; (c) brood with more than one mother (two mothers) and one father in a brood; (d) brood genetically unrelated to the brooder male; and (e) brood with more than one father (two fathers) and one mother in a brood.

G. aprion females were reported to range from 104 to 532 eggs [17,18]. As seen in aquaria, females may also approach brooding males and try to mate with them [22]. Brooding *S. nematoptera* males, a confamilial of *G. aprion*, were reported to receive a new egg clutch from another female in the wild [7]. Multiple mating by males may be a consequence of females being attracted to males that are already mouthbrooding [40] or a strategy to replenish the brooder's energy reserves to maintain condition while brooding [41].

We uncovered novel insights into the poorly known mating behaviour of paternal mouthbrooding fishes in the wild. While *N. graeffei* and *G. aprion* broods were largely monogamous with the brooder male as the sire, *G. aprion* surprisingly exhibited some degree of heterogeneity in its mating pattern—a potential reflection of alternative reproductive behaviours that aim to increase reproductive success. We highlight the potential for complex interrelationships of male care, paternity confidence and mating behaviour in mouthbrooding fishes, and suggest caution in generalized assumptions of parentage of animals displaying parental care.

Ethics. This research project was approved by the Charles Darwin University Animal Ethics Committee (permit no. A16032; A19022).

Data accessibility. Data and codes are available in the Dryad Digital Repository: <https://doi.org/10.5061/dryad.905qftmm> [42].

The data are provided in the electronic supplementary material [43].

Authors' contributions. J.E.A.: conceptualization, formal analysis, investigation, methodology, project administration, validation, visualization, writing—original draft and writing—review and editing; A.J.K.: conceptualization, funding acquisition, project administration, resources, supervision, validation, writing—original draft and writing—review and editing; O.J.L.: conceptualization, supervision, validation and writing—review and editing; D.A.C.: conceptualization, supervision, validation and writing—review and editing; D.W.: investigation, methodology, resources, supervision and writing—original draft; S.C.B.: conceptualization, formal analysis, investigation, project administration, resources, software, supervision, validation, writing—original draft and writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. This work was supported by Australian Research Council (grant no. LP150100388) (A.J.K.), Northern Territory Government and CDU RTP Scholarship and Advancement team grant (J.E.A.).

Acknowledgements. We acknowledge initial conversations with Jane Hughes and the Traditional Owners of lands and waters in which this research was conducted.

References

- Keenleyside MH. 1991 *Cichlid fishes: behaviour, ecology and evolution*. New York, NY: Springer Science & Business Media (Van Nostrand Reinhold).
- Oppenheimer JR. 1970 Mouthbreeding in fishes. *Anim. Behav.* **18**, 493–503. (doi:10.1016/0003-3472(70)90045-X)
- Gross MR, Sargent RC. 1985 The evolution of male and female parental care in fishes. *Am. Zool.* **25**, 807–822. (doi:10.1093/icb/25.3.807)
- Ostlund-Nilsson S, Nilsson GE. 2004 Breathing with a mouth full of eggs: respiratory consequences of mouthbrooding in cardinalfish. *Proc. R. Soc. B* **271**, 1015–1022. (doi:10.1098/rspb.2004.2700)
- Coleman SW, Jones AG. 2011 Patterns of multiple paternity and maternity in fishes. *Biol. J. Linn. Soc.* **103**, 735–760. (doi:10.1111/j.1095-8312.2011.01673.x)

6. Verba JT, Rabello Neto JG, Zuanon J, Farias I. 2014 Evidence of multiple paternity and cooperative parental care in the so called monogamous silver arowana *Osteoglossum bicirrhosum* (Osteoglossiformes: Osteoglossidae). *Neotrop. Ichthyol.* **12**, 145–151. (doi:10.1590/S1679-62252014000100015)
7. Rueger T, Harrison HB, Gardiner NM, Berumen ML, Jones GP. 2019 Extra-pair mating in a socially monogamous and paternal mouth-brooding cardinalfish. *Mol. Ecol.* **28**, 2625–2635. (doi:10.1111/mec.15103)
8. DeWoody JA, Avise JC. 2001 Genetic perspectives on the natural history of fish mating systems. *J. Hered.* **92**, 167–172. (doi:10.1093/jhered/92.2.167)
9. Avise JC, Jones AG, Walker D, DeWoody JA collaborators. 2002 Genetic mating systems and reproductive natural histories of fishes: lessons for ecology and evolution. *Annu. Rev. Genet.* **36**, 19–45. (doi:10.1146/annurev.genet.36.030602.090831)
10. Benun-Sutton F, Wilson AB. 2019 Where are all the moms? External fertilization predicts the rise of male parental care in bony fishes. *Evolution* **73**, 2451–2460. (doi:10.1111/evo.13846)
11. Gross MR. 2005 The evolution of parental care. *Q. Rev. Biol.* **80**, 37–45. (doi:10.1086/431023)
12. Ah-King M, Kvarnemo C, Tullberg BS. 2005 The influence of territoriality and mating system on the evolution of male care: a phylogenetic study on fish. *J. Evol. Biol.* **18**, 371–382. (doi:10.1111/j.1420-9101.2004.00823.x)
13. Gross MR. 1996 Alternative reproductive strategies and tactics: diversity within sexes. *Trends Ecol. Evol.* **11**, 92–98. (doi:10.1016/0169-5347(96)81050-0)
14. DeWoody JA, Fletcher DE, Mackiewicz M, Wilkins SD, Avise JC. 2000 The genetic mating system of spotted sunfish (*Lepomis punctatus*): mate numbers and the influence of male reproductive parasites. *Mol. Ecol.* **9**, 2119–2128. (doi:10.1046/j.1365-294X.2000.01123.x)
15. Sefc KM, Hermann CM, Taborsky B, Koblmüller S. 2012 Brood mixing and reduced polyandry in a maternally mouthbrooding cichlid with elevated among-breeder relatedness. *Mol. Ecol.* **21**, 2805–2815. (doi:10.1111/j.1365-294X.2012.05573.x)
16. Bishop K, Allen S, Pollard D, Cook M. 2001 *Ecological studies on the freshwater fishes of the Alligator Rivers Region, Northern Territory: autoecology*. Supervising Scientist report 145. Darwin, Australia: Supervising Scientist, Environment Australia. See <https://www.awe.gov.au/sites/default/files/documents/ssr145.pdf>.
17. Abecia JE, Luiz OJ, Crook DC, Banks SC, Wedd DJ, King AJ. 2021 Morphological changes and reproductive costs in brooders of two mouthbrooding freshwater fishes. *Ecol. Freshw. Fish.* **31**, 369–378. (doi:10.1111/eff.12636)
18. Pusey B, Kennard M, Arthington A. 2004 *Freshwater fishes of north-eastern Australia*. Victoria, Australia: CSIRO publishing.
19. Abecia JE, Luiz OJ, King AJ. 2018 Intraspecific morphological and reproductive trait variation in mouth almighty *Glossamia aprion* (Apogonidae) across different flow environments. *J. Fish Biol.* **93**, 961–971. (doi:10.1111/jfb.13821)
20. King AJ, Doidge C, Buckle D, Tyler KJ. 2019 Preliminary evidence of spawning phenologies of freshwater fish in a wet–dry tropical river: the importance of both wet and dry seasons. *Mar. Freshw. Res.* **71**, 202–212. (doi:10.1071/MF18458)
21. Tyler KJ, Wedd D, Crook DA, Kennard MJ, King AJ. 2021 Hydrology drives variation in spawning phenologies and diversity of larval assemblages of Australian wet–dry tropical fish. *Freshw. Biol.* **66**, 1949–1967. (doi:10.1111/fwb.13802)
22. Fauth S. 2017 Husbandry and captive breeding of mouth almighty *Glossamia aprion* (Apogonidae). *Fish. Sahul.* **31**, 1118–1122.
23. Pusey BJ, Burrows DW, Kennard MJ, Perna CN, Unmack PJ, Allsop Q, Hammer MP. 2017 Freshwater fishes of northern Australia. *Zootaxa* **4253**, 1–104. (doi:10.11646/zootaxa.4253.1.1)
24. Kilian A, Wenzl P, Huttner E, Carling J, Xia L, Blois H, Aschenbrenner-Kilian M. 2012 Diversity arrays technology: a generic genome profiling technology on open platforms. In *Data production and analysis in population genomics*, pp. 67–89. Totowa, NJ: Humana Press.
25. Jombart T, Ahmed I. 2011 adegenet 1.3-1: New tools for the analysis of genome-wide SNP data. *Bioinformatics* **27**, 3070–3071. (doi:10.1093/bioinformatics/btr521)
26. Gruber B, Unmack PJ, Berry OF, Georges A. 2018 darr: An R package to facilitate analysis of SNP data generated from reduced representation genome sequencing. *Mol. Ecol. Resour.* **18**, 691–699. (doi:10.1111/1755-0998.12745)
27. R Core Team. 2021 *R: a language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. See <https://www.R-project.org/>.
28. Sievert C. 2020 *Interactive web-based data visualization with R, plotly, and shiny*. London, UK: Chapman and Hall. See <https://plotly-r.com>.
29. Huisman J. 2017 Pedigree reconstruction from SNP data: parentage assignment, sibship clustering and beyond. *Mol. Ecol. Resour.* **17**, 1009–1024. (doi:10.1111/1755-0998.12665)
30. Jones OR, Wang J. 2010 COLONY: a program for parentage and sibship inference from multilocus genotype data. *Mol. Ecol. Resour.* **10**, 551–555. (doi:10.1111/j.1755-0998.2009.02787.x)
31. Balshine-Earn S. 1995 The costs of parental care in Galilee St Peter's fish, *Sarotherodon galilaeus*. *Anim. Behav.* **50**, 1–7. (doi:10.1006/anbe.1995.0214)
32. Okuda N. 2001 The costs of reproduction to males and females of a paternal mouthbrooding cardinalfish *Apogon notatus*. *J. Fish Biol.* **58**, 776–787. (doi:10.1111/j.1095-8649.2001.tb00530.x)
33. Alonzo SH, Klug H. 2012 Paternity, maternity, and parental care. In *The evolution of parental care* (eds NJ Royle, PT Smiseth, M Kölliker), pp. 189–205. Oxford, UK: Oxford University Press.
34. Westneat DF, Sherman PW. 1993 Parentage and the evolution of parental behavior. *Behav. Ecol.* **4**, 66–77. (doi:10.1093/beheco/4.1.66)
35. Klug H. 2018 Why monogamy? A review of potential ultimate drivers. *Front. Ecol. Evol.* **6**, 30. (doi:10.3389/fevo.2018.00030)
36. Whiteman EA, Coté IM. 2004 Monogamy in marine fishes. *Biol. Rev.* **79**, 351–375. (doi:10.1017/S1464793103006304)
37. Okuda N, Tayasu I, Yanagisawa Y. 1998 Determinate growth in a paternal mouthbrooding fish whose reproductive success is limited by buccal capacity. *Evol. Ecol.* **12**, 681–699. (doi:10.1023/A:1006533531952)
38. Hess HC. 1993 Male mouth brooding in jawfishes (Opistognathidae): constraints on polygyny. *Bull. Mar. Sci.* **52**, 806–818.
39. Sefc KM, Hermann CM, Koblmüller S. 2009 Mating system variability in a mouthbrooding cichlid fish from a tropical lake. *Mol. Ecol.* **18**, 3508–3517. (doi:10.1111/j.1365-294X.2009.04295.x)
40. Goldberg RL, Downing PA, Griffin AS, Green JP. 2020 The costs and benefits of paternal care in fish: a meta-analysis. *Proc. R. Soc. B* **287**, 20201759. (doi:10.1098/rspb.2020.1759)
41. Takeyama T, Okuda N, Yanagisawa Y. 2007 Filial cannibalism as a conditional strategy in males of a paternal mouthbrooding fish. *Evol. Ecol.* **21**, 109–119. (doi:10.1007/s10682-006-9127-7)
42. Abecia JE, King AJ, Luiz OJ, Cook DA, Wedd D, Banks SC. 2022 Data from: diverse parentage relationships in paternal mouthbrooding fishes. Dryad Digital Repository. (doi:10.5061/dryad.905qftmn)
43. Abecia JE, King AJ, Luiz OJ, Crook DA, Wedd D, Banks SC. 2022 Diverse parentage relationships in paternal mouthbrooding fishes. FigShare. (<https://doi.org/10.6084/m9.figshare.c.5958835>)