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UCT943, a next generation *Plasmodium falciparum* PI4K inhibitor preclinical candidate for the treatment of malaria

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Abstract
The 2-aminopyridine MMV048 was the first drug candidate inhibiting *Plasmodium* phosphatidylinositol 4-kinase (PI4K), a novel drug target for malaria, to enter clinical development. In an effort to identify the next generation of PI4K inhibitors, the series was optimized to improve properties such as solubility and antiplasmodial potency across the parasite lifecycle, leading to the 2-aminopyrazine UCT943. The compound displayed higher...
asexual blood stage, transmission-blocking, and liver stage activity than MMV048 and was more potent against resistant *P. falciparum* and *P. vivax* clinical isolates. Excellent *in vitro* antiplasmodial activity translated into high efficacy in *P. berghei* and humanized *P. falciparum* NOD-scid IL-2Rγnull mouse models. The high passive permeability and high aqueous solubility of UCT943, combined with low to moderate *in vitro* intrinsic clearance, resulted in sustained exposure and high bioavailability in preclinical species. In addition, the predicted human dose for a curative single administration using monkey and dog pharmacokinetics was low, ranging from 50 to 80 mg. As a next generation *Plasmodium* PI4K inhibitor, the combined preclinical data suggest that UCT943 has the potential to form part of a single-exposure radical cure and prophylaxis (SERCaP) to treat, prevent and block the transmission of malaria.

1 INTRODUCTION

Malaria, an infectious disease transmitted to people through the bite of female *Anopheles* mosquitoes infected with *Plasmodium falciparum* (*Pf*) or *Plasmodium vivax* (*Pv*), still afflicts millions of people, with almost 90% of cases on the African continent. Even though the number of malaria cases has fallen globally from an estimated 237 million cases in 2010 to 216 million cases in 2016, malaria still causes 445 000 deaths per year, 99% of which are due to *Pf* in Africa (1). Parasite resistance against currently recommended artemisinin-based combination therapies (2, 3) is a concern, and antimalarial drugs with a novel mode of action are urgently needed. We previously reported the identification of the 2-aminopyridine compound MMV390048 (also known as MMV048), which is efficacious *in vivo* against all measurable *Plasmodium* life cycle stages, except hypnozoites (4). MMV048
acts through the inhibition of *Plasmodium* phosphatidylinositol 4-kinase (PI4K) and is the first and sole agent with this mode of action that has entered clinical development. PI4K was reported as a target for *Plasmodium* in 2013 with inhibition by other compound classes (5). The low aqueous solubility associated with MMV048 in biorelevant media was identified as one of the issues to address in the next generation of PI4K inhibitors along with improved potency. Towards this goal, a scaffold change from the 2-aminopyridine to the 2-aminopyrazine core with concomitant introduction of aqueous solubilizing groups delivered analogues with a better developability profile with respect to improved physicochemical properties, as well as a significant improvement in potency across the parasite lifecycle.

Improved aqueous solubility was optimally achieved through the incorporation of a piperazinylamide group on the phenyl ring at the 5-position of the 2-aminopyrazine scaffold, leading to UCT943 (Figure 1). This compound, among other attributes, showed potent *in vitro* activity against multiple stages of the parasite lifecycle and excellent *in vivo* efficacy in the *Plasmodium berghei* and *P. falciparum* NSG (NOD-scid IL-2Rγnull) mouse models (6). In order to assess the potential of UCT943 as a follow-on compound to MMV048 and a preclinical antimalarial candidate, physicochemical, parasitological, and pharmacological profiling was undertaken. Furthermore, extensive drug metabolism and pharmacokinetics (DMPK) profiling was carried out in order to facilitate the prediction of human pharmacokinetic (PK) parameters and the efficacious single dose in humans. The results are reported herein.

2 MATERIAL AND METHODS
2.1 Chemistry

UCT943 was synthesized in seven steps from commercially available 2-aminopyrazine, as previously described (6).

2.2 In vitro antimalarial activity

2.2.1 Asexual blood stage assays

2.2.1.1 Cross-resistance against field isolates

UCT943 was tested using the [3H]-hypoxanthine incorporation assay (7, 8) against a panel of drug sensitive and drug resistant *P. falciparum* strains (Supplementary Material Table S1), as well as against a panel of resistant *P. falciparum* clones generated in the laboratory of Prof David Fidock (Columbia University, USA) (Supplementary Material Table S2).

2.2.1.2 In vitro *P. falciparum* resistance generation to UCT943 and pi4k gene sequencing

The generation of UCT943-resistant *Pf* clones was performed as described elsewhere (4) (see Supplementary Material Table S4).

2.2.1.3 Ex vivo assay against resistant *P. falciparum* clinical isolates from Côte d’Ivoire

Drug susceptibility of *P. falciparum* isolates from Côte d’Ivoire, West Africa was measured using incorporation of SYBR® Green into the parasite’s DNA as described before (9). The drug plates contained 10 serial concentrations of the antimalarials, with maximum concentration of 1170 nM for UCT943.

2.2.1.4 Ex Vivo schizont maturation drug susceptibility assay against *P. vivax* and *P. falciparum* clinical isolates

Drug susceptibility of *P. vivax* and *P. falciparum* isolates from Papua, Indonesia was measured using a modified schizont maturation assay as described previously (9). The drug plates contained 11 serial concentrations of the antimalarials, with maximum
concentrations of 2993 nM for chloroquine, 1029 nM for piperaquine, 338 nM for mefloquine, 49 nM for artemisin, and 297 nM for UCT943.

2.2.2 Liver stage assays

2.2.2.1 P. berghei liver stage assay

*Plasmodium berghei* luciferase sporozoites were obtained by dissection of infected *Anopheles stephensi* mosquito salivary glands. The sporozoite invasion assay was performed as described in (6) using the rodent parasite *P. berghei* that is able to infect human hepatocarcinoma HepG2-A16-CD81EGFP cells (10, 11).

2.2.2.2 P. cynomolgi liver stage assay

Primary rhesus hepatocytes were infected *in vitro* with *P. cynomolgi* sporozoites and the drug assays performed as previously reported by Zeeman et al. (12).

2.2.2.3 P. vivax liver stage assay

The *P. vivax* liver stage assay was implemented in human hepatocytes, infected *in vitro* with *P. vivax* sporozoites, according to the protocol described in (13).

2.2.3 Gametocyte assays

*In vitro* gametocytocidal activity was determined using luciferase reporter lines specifically enabling screening against early stage gametocytes (>90% stage I-III) and late stage gametocytes (>95% stage IV-V) as per Reader et al. (14). Methylene blue (5 μM) and MMV048 (5 μM) were routinely included as controls.

2.2.4 *P. falciparum* Dual Gamete Formation Assay (Pf DGFA)

Transmission-blocking activity of UCT943 was assessed in the DGFA, which utilizes a dual read-out that individually and simultaneously reports on the functional viability of male and female mature stage V gametocytes, as per Ruecker et al. (15).
2.3 Physicochemical properties

The pKa of UCT943 was determined by potentiometric titration as described previously (16). Solubility was measured after 24 h incubation of solid material with media at 37°C with residual solids checked by XRPD. Media included five pH buffers (pH 2.0, 4.0, 6.0, 8.0, and 10.0) and three bio-relevant media: Simulated Gastric Fluid (SGF) pH 1.8, Fasted State Simulated Intestinal Fluid (FaSSIF) pH 6.5 and Fed State Simulated Intestinal Fluid (FeSSIF) pH 5.0. Analyses were done by High Performance Liquid Chromatography (HPLC) (Waters Xbridge C18, 150 × 4.6 mm, 3 µm) at 40°C with a mobile phase of 0.1% trifluoroacetic acid (TFA) in water and 0.1% TFA in acetonitrile with UV detection (220 nm, reference 500 nm).

2.4 In vitro metabolism studies

Metabolic stability of UCT943 (1 μM) was assessed in human, dog, rat, and mouse liver microsomes using a 5-point assay and LC-MS/MS as described in (17). Metabolic stability (1 μM) was also evaluated with cryopreserved hepatocytes from the same species (1 x 10^6 viable cells/mL), as described in (18). Hepatic extraction ratio’s (E_H) were calculated using physiological based scaling factors as previously described (19).

Binding to plasma proteins and microsomal proteins (0.5 mg/mL) was determined by ultracentrifugation with LC-MS analysis as described in (20) and (17), respectively.

Permeability was determined across Caco-2 monolayers in both apical to basolateral and basolateral to apical directions using pH 7.4 in both apical and basolateral chambers, as reported in (9).

Cytochrome P450 (CYP450) inhibition studies (CYP2D6, CYP2C9, CYP3A4/5) were carried out with pooled human liver microsomes using the conditions described in (21). Metabolite identification was performed by LC-MS/MS, as described in (22), with a Phenomenex...
Kinetex PFP column, 2.1 mm x 100 mm, 2.6 μm particles using microsomal incubations, incubations in hepatocytes from the hepatocyte stability assay, and in vivo mouse PK samples.

Plasma stability and whole blood-to-plasma partitioning ratio (B:P) were determined by spiking blood from humans (Australian Red Cross Blood bank), dogs, rats, or mice with UCT943 and incubating for 4 h at 37°C. During the incubation period, aliquots of blood were taken to confirm stability. At the end of the incubation period, duplicate aliquots of blood were taken and the remaining sample was centrifuged to collect duplicate aliquots of plasma. Concentrations in blood and the plasma fraction of blood were measured by LC-MS and the blood to plasma concentration ratio calculated using the mean concentration for each matrix.

2.5 Ethics statement

Animal experiments were approved by the institutional animal care and use committees for each of the experimental sites. All studies were conducted according to the appropriate legislation and respective institutional policies on animal use and welfare.

The human biological samples were sourced ethically and their research use was in accord with the terms of the informed consents.

2.6 Pharmacokinetic studies

Pharmacokinetic studies were performed in mice, rats, dogs and monkeys as described in the Supplementary Material. For comparison of blood clearance (CL_b) to hepatic blood flow, values of 90, 55, 31, 44, and 21 mL/min/kg were assumed in mice, rats, dogs, monkeys, and humans, respectively (23).
2.7 In vivo efficacy studies

In vivo efficacy studies were conducted in the *P. berghei* and in the *P. falciparum* NOD-scid IL-2Rγnull (NSG) model as described previously (6, 24) (see Supplementary Material).

2.8 Prediction of human pharmacokinetics and efficacious single dose by pharmacokinetic/pharmacodynamic (PK/PD) modeling

2.8.1 Allometric scaling

The calculation of human plasma clearance (CLp) and human plasma volume of distribution at steady state (Vss) for UCT943 was carried out by a hybrid approach to allometric inter-species scaling (mouse, rat, dog, and monkey) of *in vivo* plasma CLp and Vss (25). After logarithmic/logarithmic transformation, the parameters were fitted to the equation \( \log y = \log a + b \log BW \), where BW is body weight; a and b are the allometric coefficient and exponent, respectively. Body weights of 0.025; 0.3; 5; 10 and 70 kg were used for mice, rats, monkeys, dogs, and humans, respectively. The mean residence time (MRT) was calculated from the predicted human plasma Vss divided by the predicted human plasma CLp.

2.8.2 PK/PD analysis in the Pf-infected NSG mouse model to predict minimum parasiticidal concentration (MPC)

Blood PK data from Pf-infected NSG mice were first fitted to a one-compartment model with first-order absorption and elimination. The predicted PK profiles were used to run a direct effect (DE) PK/PD model using Phoenix WinNonlin® (Certara, Princeton, NJ) in order to determine the minimum parasiticidal concentration (MPC) of UCT943 (Supplementary Material Equation S1 and S2).
2.8.3 Simulation of human PK profiles for human dose prediction

Dog and monkey data were used to predict human PK, since higher species allow more extensive blood sampling to explore blood or plasma concentration against time curves in detail. Intravenous (IV) and oral (PO) time course profiles were normalized in PKSolver (Excel) for preclinical species and Wajima-transformed (26). The absorption rate constant ($k_a$), and the bioavailability ($F$) estimates were obtained from PKSolver using the Wajima-transformed data. Human PO PK parameters were predicted in Berkeley Madonna (University of California, Berkeley, CA) using the Wajima transformed PK data, the human PK parameters obtained from allometry in section 2.8.1, and the $K_a$ and $F$ estimates, to have drug concentration above the MPC for ≥ 8 days. The single dose required to maintain the human plasma concentration above the MPC for eight days (section 2.8.2) was determined through simulation using the human PO PK profiles with Berkeley Madonna.

2.9 In vitro cytotoxicity, cardiotoxicity and genotoxicity

2.9.1 Cytotoxicity

In vitro cytotoxicity of UCT943 was tested against L6 cells using the Alamar Blue assay, and against Chinese Hamster Ovarian (CHO), Vero, and HepG2 cells, using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) assay (27, 28).

2.9.2 Cardiotoxicity

UCT943 was tested for inhibition of the human ether a go-go related gene (hERG) $K^+$ channel ($K_{1.1}$), human $K_{1.5} K^+$ channel, and human voltage-gated sodium channel $Na_{1.5}$ using IonWorks patch clamp electrophysiology (29), and for inhibition of the human $Ca_{1.2}$ calcium channel (cardiac $L$-type) using a fluorescence $Ca^{2+}$ assay. For all assays, 50%
cytotoxic concentrations (CC50) were determined from 8-point dose-response curves generated using 3-fold serial dilutions from the maximum final assay concentration.

2.9.3 Genotoxicity

2.9.3.1 Ames

UCT943 was assessed for mutagenic toxicity by measuring its ability to induce reverse mutations in the *Salmonella typhimurium-Escherichia coli* microsome plate incorporation assay (30, 31). Maximum concentrations were 5000 μg per plate.

2.9.3.2 Micronucleus

UCT943 capability to induce clastogenicity/aneugenicity in CHO-WBL cells was determined by measuring the extent of micronucleus formation with and without exogenous metabolic activation (Aroclor 1254 induced rat liver S9). Maximum concentrations were 500 μg/mL.

2.10 In vivo glucose-6-phosphate dehydrogenase (G6PD)-hemolysis

*In vivo* hemolytic toxicity was assessed in NOD-scid mice engrafted with A-G6PD-deficient human red blood cells (huRBCs) as described in (32) following 4-day treatment regimen at 1.5 and 10 mg/kg/day dose levels given orally.

3 RESULTS AND DISCUSSION

3.1 Rationale for optimization of MMV048 resulting in UCT943

Although MMV048 showed good potency against asexual blood stage parasites, there was room for improvement with respect to activity against liver and the transmissible gametocyte stage parasites (4).

During First-In-Human (FIH) studies, MMV048 showed high variability in exposure, which was attributed to low solubility. Intensive and time consuming formulation work had to be
carried out in order to identify a new formulation with a more consistent and 3-fold greater exposure (www.mmv.org/newsroom/interviews/mmv048-0).

To avoid such developability issues in future, improved solubility, pH-dependent and in biorelevant media, was identified as the main differentiating factor in follow-on compounds. Thus chemical modifications to MMV048 were made in such a way as to incorporate solubility-enhancing moieties. In this regard, the methyl sulfonyl group of MMV048 was replaced by a water solubilizing piperazinyl carboxamide on the phenyl ring at the 5-position of the 2-aminopyrazine core to deliver UCT943 (6). Improvements in asexual blood stage and liver stage activities were achieved by replacing the 2-aminopyridine ring with a 2-amino pyrazine ring, which also maintained good potency against gametocytes (6). One key objective of this study was firstly to determine if these improvements would translate into better in vivo efficacy in the Pf-infected NSG mouse model and accordingly, a low predicated human dose.

3.2 In vitro antimalarial activity

The biological target of the clinical candidate MMV048 was identified to be Plasmodium phosphatidylinositol 4-kinase (Pi4K) through resistant mutant generation, sequencing, and pull-down experiments (4). Plasmodium Pi4K has recently been identified as a new and promising drug target, which is present at all life-cycle stages of the Pf and Pv parasites (5). UCT943 inhibits the PvPi4K enzyme with an IC_{50} = 23 nM. When tested against a 5-fold resistant P. falciparum strain generated against MMV048 due to the mutated pfpi4k locus (4), UCT943 displayed a 6-fold shift in IC_{50} relative to the parental Dd2 strain (14 nM relative to 2.2 nM) (Supplementary Material Table S2). When resistance was selected for in Dd2-B2 using UCT943, pfpi4k mutants with a 4- or 8-fold IC_{50} shift were obtained (Supplementary...
and are located in the same region of pi4k as MMV048-selected mutations reported elsewhere (4). Resistance selection studies indicated a minimum required inoculum of $10^7$ Dd2 parasites for resistance to emerge, which is similar to MMV048 (4). By comparison, IC$_{50}$ shifts were within 2-fold when tested against strains containing mutated loci in either pf cytB, pf dhodh, pf atp4, or pf carl. These data indicate that UCT943 and MMV048 have the same molecular target, and that similarly to MMV048, UCT943 is expected to have transmission blocking and liver stage activity. Importantly, UCT943 maintained high in vitro selectivity (> 200 fold) for the parasite P$v$PI4K versus the human PI4KB isozyme (IC$_{50}$ (PI4KB) = 5.4 μM), which inhibition is linked to immunosuppressive effects (33).

UCT943 was one of the most potent compounds assessed in the 2-aminopyrazine chemical series (6), with IC$_{50}$’s of 5.4 and 4.7 nM against NF54 and K1 P. falciparum strains, respectively, which was 5- to 6-fold more active than the clinical candidate MMV048 (4) (Figure 2). In addition, UCT943 was equally active against the drug sensitive NF54 strain and multi-drug resistant strains, with IC$_{50}$’s ranging from 4 to 7 nM, thus suggesting that cross-resistance with existing antimalarials is a low risk (Supplementary Material Table S1).

UCT943 primarily exhibited blood stage activity against schizonts (Supplementary Material Table S5), which correlates with a slow rate of kill as determined in the in vitro parasite reduction ratio (PRR) assay (lag phase = 48 h, Log PRR = 2.5 at 10 x EC$_{50}$ in 3D7 strain) and in the in vitro speed assay (Supplementary Material Table S5). As expected, the killing rate profile of UCT943 was similar to that of MMV048 (lag phase = 48 h, Log PRR = 2.7) (4). When tested against Pf clinical isolates of the Ivory Coast, UCT943 exhibited potent activity (2-15 nM) (Supplementary Material Table S6). The potency of UCT943 against clinical isolates
from Papua Indonesia was significantly higher than that of MMV048 in *Pv* and in *Pf* (median IC₅₀ 14 nM and 29 nM; p=0.012 versus median IC₅₀ 93 nM and 202 nM; p<0.001) (Figure 2 and Supplementary Material Table S7). Interestingly, both compounds displayed higher potency against *Pv* than *Pf*, a trend that was also observed for another PI4K inhibitor, KDU691, albeit to a lesser extent (5).

When tested against other stages of the parasite life cycle (Figure 2), UCT943 was 2-50 times more potent than MMV048 (4). UCT943 was potent against early stage (>90% stage I-III) and late stage gametocytes (>95% stage IV-V) (IC₅₀ of 134 nM and 66 nM, respectively) and inhibited the formation of both male and female gametes (IC₅₀ ≈ 80 nM) in the dual gamete formation assay (DGFA). The latter activity translated into transmission-blocking activity (Target Candidate Profile TCP5 as defined by Burrows *et al.* (34)) in the standard membrane feeding assay (SMFA), with an IC₅₀ of 96 nM (35), equivalent to MMV048 (Figure 2). In-depth clinical PK/PD investigations would be needed to determine doses that would afford coverage for both blood stage and transmission-blocking activities. A single drug with both activities would be a valuable addition to the arsenal of antimalarial medicines.

UCT943 also exhibits better *in vitro* liver stage activity than MMV048 (potentially addressing TCP3 and TCP4 (34)), targeting schizonts *in vitro* (IC₅₀ of < 100 nM and <10 nM in *P. vivax* and *P. cynomolgi*, respectively, when tested prophylactically, and 0.92 nM in *P. berghei*), as well as inhibiting the formation of hypnozoites (IC₅₀ of < 100 nM and < 10 nM in *P. vivax* and *P. cynomolgi*, respectively when tested prophylactically) (Figure 2). The parasitophorous vacuole membrane protein UIS4 becomes internalized in MMV048-treated *P. berghei* HepG2 cultures, a morphology associated with in vitro liver stage parasite clearance (4).
Though UCT943 was not evaluated for such morphological changes, its activity against schizonts offers prospects for improved prophylactic liver stage activity relative to MMV048.

3.3 Physicochemical characterization

UCT943 was stable in the solid state over a period of 18 months and was chemically stable at 20°C in solutions of DMSO, water (pH 6.2), and buffers (pH 2, pH 7.4) over 6 days. The piperazinylamide group accounts for the significantly lower measured lipophilicity of UCT943 (LogD = -0.27) relative to MMV048 (LogD = 2.6). This, combined with a pKa of 7.5, results in higher measured solubility in aqueous media compared to MMV048, across a range of physiologically relevant pH’s up to 6.5, where the compound is protonated (Table 1). The compound was also highly soluble in SGF (pH 1.8), FeSSIF (pH 5.0), and FaSSIF media (pH 6.5) (> 1.5 mg/mL), predictive of a good dissolution in the gastrointestinal tract. The pH-dependent solubility profile of UCT943 in aqueous media was typical of a weak base with higher solubility at pH below the pKa, i.e. 7.5 (Figure 3). UCT943 exhibited high permeability across Caco-2 cells in both directions (P_{app \ B>A} = 25 \times 10^{-6} \text{ cm/s}; P_{app \ A>B} = 28 \times 10^{-6} \text{ cm/s}) without appreciable efflux (efflux ratio of 0.93, Table 2). With high FaSSIF solubility and high permeability, UCT943 could be classified as a Developability Classification System (DCS) Class I compound, which bodes well for a much more favorable development pathway compared to MMV048 that was classified as DCS Class II (Figure 4) (36). The high solubility and permeability of UCT943, along with its potent anti-*Plasmodium* activity against all stages of the parasite lifecycle (see section 3.1) triggered comprehensive DMPK profiling toward determining its potential for preclinical development.
3.4 *In vitro* and *in vivo* metabolism studies

There was no evidence of chemical instability for UCT943 in human, dog, rat, or mouse blood or plasma. The compound was moderately bound to plasma proteins with little species differentiation (f_u in plasma 0.10-0.16). It also had a greater propensity than MMV048 (B:P ~1.0 across species) to bind or distribute in red blood cells (RBCs) relative to plasma (B:P varying from 1.5 to 2.3 across species). This is likely due to enhanced partitioning through the acidic phospholipid bilayer of the cell membrane due to the weakly basic piperazine moiety. The higher partitioning into RBCs possibly contributes to a small degree to its potent *in vitro* and *in vivo* anti-*Plasmodium* activity by localizing the drug where the parasite resides.

As an indicator of hepatic metabolism, UCT943 was incubated across species with hepatocytes, liver microsomes, and liver S9 fraction (Table 2). The intrinsic clearance of UCT943 in both microsomes and hepatocytes was low in human, rat, and mouse (CL_int < 11.6 µL/min/mg in microsomes and CL_int < 4 µL/min/10^6 cells in hepatocytes), while it was moderate in dog (CL_int = 28.2 µL/min/mg in microsomes and CL_int = 9 µL/min/10^6 cells in hepatocytes). Additionally, no significant metabolism (< 10% degradation) was detected in human liver S9 fraction, showing that enzymes other than cytochrome P450s (CYPs) were not extensively involved in the metabolism of UCT943. No measurable inhibition was detected at 20 µM against any of the CYP isoforms tested (CYP2D6, CYP2C9, CYP3A4/5), indicating that UCT943 has low potential for *in vivo* enzyme inhibition and that adverse drug-drug interactions through oxidative metabolism are likely to be minimal.

Metabolite identification was performed *in vitro* using liver microsomes and hepatocytes, as well as *in vivo* by analyzing the collected blood samples from mouse PK experiments (Figure...
In vitro, liver microsomes and hepatocytes gave a complementary picture, both showing the major biotransformation pathway occurring on the piperazinylamide moiety (Figure 5b). UCT943 was mainly metabolized into an oxidation metabolite (P+16), which was further dehydrogenated into two metabolites P+14 (I&II) in non-human species. Further biotransformation on the piperazine ring gave metabolites P-26 (piperazine ring cleavage) and P+28. The formation of the carboxylic acid derivative by hydrolysis of the piperazinylamide moiety (P-68) was more easily detected in hepatocytes and in vivo in mice than in liver microsomes, presumably due to the higher concentration of the enzymes responsible for this particular biotransformation. The contribution of these metabolites to the activity of the parent UCT943 is currently under investigation. Notably, the P-68 metabolite, resulting from the hydrolysis of the carboxamide, showed high activity, albeit lower than that of the parent, with IC$_{50}$’s of 33 nM and 32 nM against *P. falciparum* NF54 and K1 strains respectively (6).

3.5 Pharmacokinetic studies
When administered intravenously, the blood clearance (CL$_b$) of UCT943 was low in mice, rats, and dogs (Table 3), with a value of less than 20% of hepatic blood flow (23) to very low in monkeys with blood CL < 5% hepatic blood flow, which is consistent with the high in vitro metabolic stability. Plasma volume of distribution was high in all species (V$_{ss}$ 7.1 - 13.1 L/kg), suggesting that the compound extensively distributes and accumulates in organ tissues. This would be expected for a basic compound due to partitioning into cell membranes by associating with acidic phospholipids (37). As a consequence of the low CL and high V$_{ss}$, half-lives were long, ranging from 6 h in mice to 29 h in monkeys. When dosed orally in mice and dogs, UCT943 was rapidly absorbed (time of maximum concentration T$_{max}$ < 3 h), while
absorption in rats and monkeys was slower with a maximum concentration (C<sub>max</sub>) reached after 12 h and 7 h, respectively (Figure 6). Oral bioavailability was high across all species, ranging from 66% to 98%. The good oral bioavailability and the long t<sub>1/2</sub> of UCT943 across species is encouraging toward achieving a single dose treatment and cure for malaria, which would boost patient compliance in resource-limited regions of the world, where the medical infrastructure is not sufficient. If these PK properties are confirmed in humans, UCT943 could thus be a potential combination partner in a single exposure radical cure and prophylaxis (SERCaP) treatment as proposed by the Medicines for Malaria Venture (MMV) (34).

3.6 In vivo efficacy studies

When dosed at 10 mg/kg p.o., UCT943 reduced parasitemia by > 99.9% in the mouse <i>P. berghei</i> infection model and cured all mice with > 30 mean survival days (MSD). At 3 mg/kg p.o., no complete cure was achieved and MSD was 10 days (6), albeit parasitemia was reduced by 99%. The resulting 90% effective dose (ED<sub>90</sub>) was 1.0 mg/kg p.o. in the <i>P. berghei</i> infection model. In the <i>Pf</i>-infected NSG mouse model (Supplementary Material Figure S1), UCT943 was 2-fold more potent than MMV048 with a ED<sub>90</sub> (90% effective dose) of 0.25 mg/kg compared to 0.57 mg/kg (4). PK data showed that the exposure of UCT943 was dose-dependent (6). As PK/PD relationships in the humanized NSG mouse model have been found to be predictive of the induced blood-stage malaria (IBSM) model in human volunteers (38), we used the NSG mouse efficacy data for human dose prediction (see next section 3.7).
3.7 Prediction of human pharmacokinetics and efficacious single dose by PKPD modeling

3.7.1 Allometric scaling

The slope used to predict plasma CL by allometric scaling was close to 0.75 (0.73), while the slope used to predict plasma Vss was close to 1 (0.90), as expected for metabolic processes and volumes, respectively (39) (Figure 7). The predicted human plasma Vss was moderately large (6.3 L/kg) and the predicted human plasma CL was low (0.20 L/h/kg) as shown in Table 4. When converted using the B:P ratio of 1.5, the predicted human blood clearance was 0.10 L/h/kg, i.e. less than 10% of liver blood flow (23). The predicted t1/2 in humans was 27 h, which together with a long mean residence time of 32 h, suggests that the compound will be a long-duration antimalarial agent.

3.7.2 Estimation of minimum parasiticidal concentration (MPC) in blood in the Pf-infected NSG mouse model

From the PK/PD model (Supplementary Material Figure S2 and S3), the compound specific PD parameters EC50 and Kkill were estimated to be 3.7 ng/mL and 0.060 /h respectively. The MPC in blood (MPCb), calculated using the EC50, was 7.4 ng/mL in NSG mouse blood and 5.2 ng/mL in human blood (Table 4), after correction for Pf-infected NSG mouse and human B:P partitioning data (2.8 and 1.5, respectively). The MPC in plasma (MPCp) used for human dose prediction was therefore 2.6 ng/mL. The in vivo parasite reduction ratio (PRR) of 48 h predicted by the PK/PD model (Log PRR = 1.25) correlated closely to the in vitro moderate killing profile (Log PRR = 2.5), confirming UCT943 as a slow acting antimalarial compound in this model.
3.7.3 Prediction of human pharmacokinetic parameters and efficacious dose

The human PK profiles modeled in Berkeley Madonna predicted a single human dose of 50-80 mg, based on dog and monkey data, respectively, in order to maintain the plasma concentrations above the predicted therapeutic level (that is, the plasma MPCₚ of 2.6 ng/mL) for 8 days (i.e. four asexual parasite cycles) (Supplementary Material Figure S4 and S5). In this model, a single administration of the efficacious dose to maintain plasma concentrations above the MPCₚ for 8 days resulted in an area under the curve (AUC) of 4213-8223 ng·h/mL, and a predicted Cₘₐₓ of 234-358 ng/mL, based on monkey and dog, respectively. The low predicted dose is particularly encouraging, since it leaves a generous margin for potential dose increases, in case the predicted value was underestimated, or dose increase was deemed desirable to prevent the development of resistance or to ensure activity against other malaria species (including P. vivax, see section 3.1).

3.8 In vitro cytotoxicity, cardiotoxicity and genotoxicity

Cytotoxicity, assessed against four mammalian cell lines, was found to be low with a selectivity index (SI) greater than 2200 relative to the IC₅₀ in NF54 and greater than 170-fold against the highest IC₅₀ in Pf clinical isolates (Table 5). The SI is equivalent or greater than that of MMV048. Relative to the predicted upper unbound Cₘₐₓ (0.13 μM) for the human efficacious plasma exposure, a 90-fold margin is thereby predicted and is sufficiently high to warrant progression into in vivo preclinical toxicology studies.

The safety margins over cardiotoxicity risk were largely improved compared to MMV048 (see selectivity indexes SI in Table 5). The hERG IC₅₀ of 10 μM corresponds to an 80-fold margin relative to the predicted therapeutic unbound Cₘₐₓ exposure for efficacy, which indicates a low risk of QT interval prolongation at therapeutic exposures. The margins to
potential safety issues associated with potential off-target activities at other ion channels (NaV1.5, CaV1.2, and Kv1.5) are even higher (Table 5).

Genotoxicity was evaluated using the Ames and mouse micronucleus tests, in which UCT943 tested negative at the highest concentrations (Table 5), suggesting that the compound does not have the potential to result in back-mutation of a defective gene to recover its function (Ames test) (30), and does not have the ability to induce the formation of micronuclei during cell division as a consequence of genetic damage (micronucleus assay)(40).

3.9 In vivo G6PD-hemolysis

In the search for new antimalarial compounds, it is essential to develop drugs which do not pose a red blood cell hemolysis risk to patients with G6PD deficiency (34). When assessed for in vivo hemolytic toxicity in NOD-scid mice engrafted with A-G6PD-deficient human red blood cells (huRBCs), UCT943 showed comparable day 7 huRBC levels as those treated with the vehicle control, these levels being significantly higher when compared against the positive control primaquine (25 mg/kg/day). This indicates that UCT943 does not induce hemolytic toxicity at neither 1.5 nor 10 mg/kg/day dosing (Supplementary Material Figure S6), which is higher than the ED$_{90}$ of 0.25 mg/kg in the P. falciparum NSG model of infection.

Assessment of other markers of hemolysis, including spleen weight and mouse reticulocyte levels, also supports that UCT943 did not induce hemolytic toxicity (Supplementary Material Figures S7a and S7b).

4 CONCLUSION

UCT943 was optimized for antiplasmodial activity from a series of 2-aminopyrazines by structural modification of the clinical candidate MMV048, an inhibitor of an essential
Plasmodium enzyme, PI4K. Incorporation of a piperazinylamide group resulted in enhanced water solubility while maintaining high permeability, both parameters being key for a good developability profile and for achieving high drug exposure. This, combined with minimal in vitro metabolism in liver subcellular fractions and in hepatocytes, translated into low clearance, sustained exposure, and high bioavailability in preclinical species. UCT943 was potent against all stages of the Plasmodium parasite lifecycle, as well as against resistant *P. falciparum* and *P. vivax* clinical isolates. The 5-fold better in vitro antiplasmodial activity of UCT943 compared to MMV048 translated into excellent efficacy in the *P. berghei* mouse model and improved efficacy in the humanized *P. falciparum* mouse model. UCT943 was found to be a slow acting, long duration antimalarial compound similar to what is seen for quinoline antimalarials such as mefloquine. The predicted human single dose using monkey and dog pharmacokinetics was low, ranging from 50 to 80 mg, which offers considerable potential for the drug candidate. The high safety margins over cytotoxicity and cardiac toxicity highlighted herein are much larger than the predicted human therapeutic exposure, which is promising. Based on the data presented, UCT943 displays asexual blood stage, transmission-blocking, and liver stage activity and thus has the potential to form part of a single-exposure radical cure and prophylaxis (SERCaP) treatment of uncomplicated malaria. This breadth of activity offers considerable flexibility with respect to treatment options and TPPs that might be addressed and have contributed to the selection of UCT943 for preclinical development as a follow-on compound to MMV048.

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Africa) for the ADME assays; Virgil Verhoog and Sumaya Salie from H3D, University of Cape Town (South Africa) for the Pf blood stage assays; Trevor Finch from the Division of Pharmacology, University of Cape Town (South Africa) for assistance with the animal work; Michael Delves, Andrea Ruecker and Robert E. Sinden from the Cell and Molecular Biology laboratory, Imperial College, London (United Kingdom) for the gamete formation assay; Anne-Marie Zeeman and Clemens H. M Kocken from the Biomedical Primate Research Centre, Rijswijk (The Netherlands) for the Pc in vitro prophylactic and radical cure assay; Rachaneeporn Jenwithisuk from the Faculty of Tropical Medicine, Mahidol University, Bangkok (Thailand) for the Pv in vitro prophylactic and radical cure assay; John Burke from the University of Victoria, British Columbia (Canada) for the PvPI4K assay.

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**Table 1: Physico-chemical properties of UCT943 compared to MMV048**

<table>
<thead>
<tr>
<th>Property (SD)</th>
<th>UCT943</th>
<th>MMV048</th>
</tr>
</thead>
<tbody>
<tr>
<td>MW (g/mol)</td>
<td>427.4</td>
<td>393.4</td>
</tr>
<tr>
<td>LogD pH 7.4</td>
<td>-0.27 (0.01)</td>
<td>2.6 (0.03)</td>
</tr>
<tr>
<td>pKa measured</td>
<td>7.45 (0.05)</td>
<td>4.0 (0.07)</td>
</tr>
<tr>
<td>Thermodynamic solubility (μg/mL)*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>pH 2.0</td>
<td>3000</td>
<td>740</td>
</tr>
<tr>
<td>pH 4.0</td>
<td>49</td>
<td>-</td>
</tr>
<tr>
<td>pH 6.0</td>
<td>110</td>
<td>4.2 (pH 6.5)</td>
</tr>
<tr>
<td>pH 8.0</td>
<td>31</td>
<td>4.0 (pH 7.4)</td>
</tr>
<tr>
<td>pH 10.0</td>
<td>8.3</td>
<td>-</td>
</tr>
<tr>
<td>SGF (pH 1.8)</td>
<td>5900</td>
<td>-</td>
</tr>
<tr>
<td>FaSSIF (pH 6.5)</td>
<td>1500</td>
<td>14.4</td>
</tr>
<tr>
<td>FeSSIF (pH 5.0)</td>
<td>1900</td>
<td>28.3 (pH 5.8)</td>
</tr>
</tbody>
</table>

SGF: Simulated Gastric Fluid; FaSSIF: Fasted State Simulated Intestinal Fluid; FeSSIF: Fed State Simulated Intestinal Fluid

* single determination
Table 2: *In vitro metabolism, permeability, protein binding, blood:plasma ratio and plasma stability data for UCT943*

<table>
<thead>
<tr>
<th>Parameter</th>
<th>h/d/r/m (SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microsomal CL$_{int}$ (μL/min/mg)</td>
<td>&lt;11.6 (0.1) / 28.2 (0.5) / 11.6 (0.2) / &lt;11.6 (0.7)</td>
</tr>
<tr>
<td>Hepatocyte CL$_{int}$ (μL/min/10$^6$ cells)*</td>
<td>&lt;2 / 9 / 4 / &lt;4</td>
</tr>
<tr>
<td>Hepatocyte predicted E$_{H}$*</td>
<td>&lt;0.2 / 0.65 / 0.23 / &lt;0.2</td>
</tr>
<tr>
<td>Caco-2 P$<em>{app}$ B$\rightarrow$A / P$</em>{app}$ A$\rightarrow$B (10$^{-6}$ cm/s)</td>
<td>25 (5) / 28 (2)</td>
</tr>
<tr>
<td>f$_u$ microsomes</td>
<td>0.15 (0.01) / 0.50 (0.04) / 0.45 (0.05) / 0.47 (0.04)</td>
</tr>
<tr>
<td>f$_u$ plasma</td>
<td>0.17 (0.009) / 0.10 (0.007) / 0.15 (0.009) / 0.10 (0.02)</td>
</tr>
<tr>
<td>Blood:Plasma ratio</td>
<td>1.5 (0.1) / 2.3 (0.1) / 2.1 (0.2) / 1.9 (0.2)</td>
</tr>
<tr>
<td>Plasma stability (% after 240 min)*</td>
<td>97 / 95 / 97 / 102</td>
</tr>
</tbody>
</table>

* single determination
Table 3: In vivo pharmacokinetic parameters for UCT943 across mouse, rat, dog, and monkey species calculated from non-compartmental analysis (SD are given in brackets)

<table>
<thead>
<tr>
<th>Species</th>
<th>Mouse</th>
<th>Mouse</th>
<th>Rat</th>
<th>Rat</th>
<th>Dog</th>
<th>Dog</th>
<th>Monkey</th>
<th>Monkey</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dose (mg/kg)</td>
<td>5 (IV)</td>
<td>20 (PO)</td>
<td>5 (IV)</td>
<td>20 (PO)</td>
<td>2 (IV)</td>
<td>10 (PO)</td>
<td>2 (IV)</td>
<td>10 (PO)</td>
</tr>
<tr>
<td>( t_{1/2} ) (h)</td>
<td>6.4 (0.7)</td>
<td>5.7 (0)</td>
<td>7.4 (0.6)</td>
<td>5.3 (0.3)</td>
<td>13.0</td>
<td>16.1</td>
<td>28.6</td>
<td>34.6</td>
</tr>
<tr>
<td>Plasma ( V_{ss} ) (L/kg)</td>
<td>13.1* (1.1)</td>
<td>-</td>
<td>11.5* (2.1)</td>
<td>-</td>
<td>7.1 (0.9)</td>
<td>-</td>
<td>8.7 (1.0)</td>
<td>-</td>
</tr>
<tr>
<td>Blood CLb (mL/min/kg)</td>
<td>12.6 (2.4)</td>
<td>-</td>
<td>9.5 (1.6)</td>
<td>-</td>
<td>3.3* (0.6)</td>
<td>-</td>
<td>2.0 * (0.3)</td>
<td>-</td>
</tr>
<tr>
<td>Plasma CLp (mL/min/kg)</td>
<td>24.0* (4.6)</td>
<td>-</td>
<td>20.0* (3.4)</td>
<td>-</td>
<td>7.5 (1.2)</td>
<td>-</td>
<td>3.9 (0.6)</td>
<td>-</td>
</tr>
<tr>
<td>Plasma AUC0-∞ (min·µM)</td>
<td>497* (8.5)</td>
<td>1310* (202)</td>
<td>725* (109)</td>
<td>2843* (480)</td>
<td>634 (89)</td>
<td>-</td>
<td>2335</td>
<td>1192</td>
</tr>
<tr>
<td>Plasma Cmax (µM)</td>
<td>1.2* (0.2)</td>
<td>1.7* (0.3)</td>
<td>2.4* (0.2)</td>
<td>2.1* (0.7)</td>
<td>1.0 (0.1)</td>
<td>2.3 (0.4)</td>
<td>1.5 (0.03)</td>
<td>2.1 (0.4)</td>
</tr>
<tr>
<td>Tmax (h)</td>
<td>-</td>
<td>4.0 (3.6)</td>
<td>-</td>
<td>12.0</td>
<td>-</td>
<td>2.3 (1.5)</td>
<td>-</td>
<td>7.0 (1.7)</td>
</tr>
<tr>
<td>F (%)</td>
<td>-</td>
<td>66 (10)</td>
<td>-</td>
<td>98 (4.9)</td>
<td>-</td>
<td>74 (23.7)</td>
<td>-</td>
<td>80 (12.8)</td>
</tr>
</tbody>
</table>

*blood values were scaled to plasma values using B:P ratios of 1.9/2.1/2.3/2.0 for mice, rats, dogs, and monkeys, respectively
Table 4: Predicted human PK parameters for UCT943 from modeling

<table>
<thead>
<tr>
<th>Parameter</th>
<th>UCT943</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma $V_{ss}$ (L/kg)</td>
<td>6.3</td>
</tr>
<tr>
<td>Plasma $CL_p$ (L/h/kg)</td>
<td>0.20</td>
</tr>
<tr>
<td>MRT (h)</td>
<td>32</td>
</tr>
<tr>
<td>Plasma MPC$_p$ (ng/mL); blood MPC$_b$ (ng/mL);</td>
<td>2.6; 5.2</td>
</tr>
<tr>
<td>$k_a$ (/h)</td>
<td>0.25</td>
</tr>
<tr>
<td>$F$ (%)</td>
<td>80</td>
</tr>
<tr>
<td>Single dose (mg)*</td>
<td>50-80</td>
</tr>
<tr>
<td>$t_{1/2}$ (h)</td>
<td>27</td>
</tr>
<tr>
<td>Plasma AUC$_p$ (ng∙h/mL)</td>
<td>4213-8223</td>
</tr>
<tr>
<td>Plasma $C_{max,p}$ (ng/mL)</td>
<td>234-358</td>
</tr>
</tbody>
</table>

MPC: Minimum Parasiticidal Concentration

*Predicted single dose to achieve ≥8 days above MPC

$\text{b}$ blood; $\text{p}$ plasma
Table 5: Cytotoxicity, cardiotoxicity and genotoxicity data for UCT943 and MMV048

<table>
<thead>
<tr>
<th></th>
<th>MMV048</th>
<th>UCT943</th>
<th>Lowest SI</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Cytotoxicity</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CHO</td>
<td>17 (3148x)</td>
<td>298</td>
<td></td>
</tr>
<tr>
<td>Vero</td>
<td>113 (20926x)</td>
<td>1982</td>
<td></td>
</tr>
<tr>
<td>HepG2</td>
<td>&gt;10 (357x)</td>
<td>13 (2407x)</td>
<td>225</td>
</tr>
<tr>
<td>L6</td>
<td>251 (8964x)</td>
<td>12 (2222x)</td>
<td>211</td>
</tr>
<tr>
<td><strong>Cardiotoxicity</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>hERG (Kv11.1)</td>
<td>&gt;11 (393x)</td>
<td>10 [6.4-15.9]</td>
<td>177</td>
</tr>
<tr>
<td></td>
<td>(1870x)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Na&lt;sub&gt;v&lt;/sub&gt;1.5</td>
<td>100 (3571x)</td>
<td>&gt;33 (&gt;6111x)</td>
<td>&gt;579</td>
</tr>
<tr>
<td>Ca&lt;sub&gt;v&lt;/sub&gt;1.2</td>
<td>16 (571x)</td>
<td>&gt;33 (&gt;6111x)</td>
<td>&gt;579</td>
</tr>
<tr>
<td>K&lt;sub&gt;v&lt;/sub&gt;1.5</td>
<td>-</td>
<td>&gt;33 (&gt;6111x)</td>
<td>&gt;579</td>
</tr>
<tr>
<td><strong>Genotoxicity</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ames assay</td>
<td>Negative</td>
<td>Negative</td>
<td>-</td>
</tr>
<tr>
<td>Micronucleus test</td>
<td>Negative</td>
<td>Negative</td>
<td>-</td>
</tr>
</tbody>
</table>

CC<sub>50</sub>: 50% cytotoxic concentration

IC<sub>50</sub>: 50% inhibitory concentration

SI: selectivity index; SI = CC<sub>50</sub> / IC<sub>50</sub>

Lowest SI; lowest selectivity index against clinical isolates; lowest SI = CC<sub>50</sub> / max IC<sub>50</sub> Pf

Clinical isolate
Figure 1 Structure of UCT943 and MMV048

Figure 2 Antiplasmodial activity of UCT943 compared to MMV048 against different stages of the parasite life cycle (values indicate IC_{50} in nM; blue: UCT943; brown: MMV048)

a Standard Membrane Feeding Assay, indirect mode (35)

b prophylactic assay
Figure 3: Comparison of pH solubility profiles of MMV048 (dotted blue line) and UCT943 (dotted red line) calculated and measured values for MMV048 (triangle dots), and UCT943 (square dots). FaSSIF: Fasted State Simulated Intestinal Fluid; FeSSIF: Fed State Simulated Intestinal Fluid.
Figure 4: Ranking of UCT943 and MMV048 in the Developability Classification System (DCS) (36).
Figure 5a: Proposed metabolic pathway of UCT943 in microsomes (mouse, rat, human), hepatocytes and in vivo in mice
Figure 5b: Metabolite profiles of UCT943 in liver microsomes, hepatocytes, and in mice

LM: liver microsomes; Hp: hepatocytes; H: human; R: rat; M: mice; D: dog
Figure 6: Whole blood concentration vs time profiles following IV (left) and oral (right) administration of UCT943 to (a, b) non-fasted male Balb/C mice; (c, d) fasted male Sprague Dawley rats; (e, f) fasted male beagle dogs; (g, h) fasted female Cynomolgus monkeys.
Figure 7: Allometric plot for UCT943 (a) plasma clearance and (b) plasma volume of distribution

(a)

(b)
References


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