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Published in:
Applied and Environmental Microbiology

DOI:
[10.1128/AEM.00287-10](https://doi.org/10.1128/AEM.00287-10)

Published: 01/07/2010

Document Version
Peer reviewed version

[Link to publication](#)

Citation for published version (APA):

Draper, A., Mayo, M., Harrington, G., Karp, D., Yinfoo, D., Ward, L., Haslem, A., Currie, B., & Kaestli, M. (2010). Association of the melioidosis agent burkholderia pseudomallei with water parameters in rural water supplies in Northern Australia. *Applied and Environmental Microbiology*, 76(15), 5305-5307.
<https://doi.org/10.1128/AEM.00287-10>

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1 **Association of the Melioidosis Agent *Burkholderia pseudomallei* with**
2 **Water Parameters in Rural Water Supplies in Northern Australia**

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17

18 Running Title: *Burkholderia pseudomallei* in Bore Water in Australia.

19 **Abstract:**

20

21 We analysed water parameters and the occurrence of the melioidosis agent, *Burkholderia*
22 *pseudomallei* in 47 water bores in Northern Australia. *B.pseudomallei* was associated with soft,
23 acidic bore water of low salinity but high iron levels. This finding aids in identifying water
24 supplies at risk of contamination with this pathogenic bacterium.

25

26 **Text**

27 Melioidosis is a severe, emerging disease caused by the Gram-negative bacterium *Burkholderia*
28 *pseudomallei* (*B.ps*), a hydrophilic soil saprophyte endemic in Southeast Asia and northern
29 Australia (3,17,18). Melioidosis is the commonest cause of fatal community-acquired
30 bacteraemic pneumonia in northern Australia (4).

31 Melioidosis outbreaks have been attributed to contaminated unchlorinated water supplies in
32 northern Australia causing fatalities among humans and animals (5,8)(own observation). Rural
33 water bores – of which there are 5,000 around Darwin alone - are mostly unchlorinated due to
34 concerns of bore owners about taste, by-products and maintenance of chlorination. We have
35 analyzed the association of *B.ps* occurrence with environmental and physico-chemical water
36 properties in unchlorinated bore water from rural Darwin in northern Australia. This study is the
37 first report on the habitat of *B.ps* in water supplies.

38 **Water sampling.** Bore water was collected from 47 properties (188 samples) in an area of 22x13
39 km in the 2008 dry season. 25 bores (53%) were fed by carbonate rock aquifers and 22 by
40 aquifers in fractured weathered rock (6,13). 26 were resampled in the wet season (103 samples).
41 Per bore, 1L of water was collected after 1, 30 and 60 minutes of water pumping to represent
42 water from the bore head, shaft and aquifer. Water was collected from tanks linked to the bore.
43 Water filtration and *B.ps* culture was done as previously described (7). Briefly, *B.ps* was cultured
44 in modified Ashdown's-Broth and Tryptone-Soy-Broth and sub-cultured onto Ashdown's agar.
45 Colonies exhibiting *B.ps* morphology were confirmed by latex agglutination and PCR targeting
46 TTS1 (14). Water samples were tested for pH, temperature, electro-conductivity (TPS), total

47 nitrates, total iron, phosphates (Hanna-Instruments) and total hardness (AquaspeX). Water
48 samples were cultured for total coliform counts (3M).

49 **Occurrence of *B.ps*.** In the dry season, 12 of 47 bores (26%) tested positive for *B.ps*. In the wet
50 season, these 12 were revisited together with 14 bores negative for *B.ps* matched for aquifer type
51 and location. 11 of the 12 initially positive bores were again positive and 4/14 (29%) previously
52 negative bores were newly positive in the wet season.

53 Multilocus variable-number tandem repeat analysis (MLVA-4) of *B.ps* isolates (10) revealed 33
54 different MLV-4 patterns with identical or closely related genotypes also found in *B.ps* from soil
55 or clinical cases within a 50 km radius. A median of 1.5 different genotypes was found per bore
56 visit (95%CI 1-2). Isolates with identical or closely related patterns in the dry and wet season
57 were retrieved from seven of the bores indicating persistent colonization. No geographical
58 clustering of positive bores nor MLV-4 types was obvious.

59 No significant variation in bore characteristics was evident between *B.ps* positive and negative
60 bores. These included bore age, depth, aquifer type, presence of concrete casing/slab, likelihood
61 of water pooling at the bore and origin of sample (bore head, shaft, aquifer, tank). Effective bore
62 capping showed reduced *B.ps* prevalence (21% vs 44%; $p=0.21$) (Table 1).

63 We analyzed the association between the occurrence of *B.ps* and water parameters (Table/Figure
64 1). A significant association was found between the occurrence of *B.ps* and more acidic water.
65 These results support data showing the preference of *B.ps* for more acidic soil (2,10,15).

66 Water hardness, i.e. calcium carbonate levels, salinity and pH showed a significant positive
67 correlation with each other (Spearman's correlation, $P<0.001$), attributed to the buffering
68 capacity of carbonates and salts. *B.ps* was significantly associated with low hardness, i.e. soft

69 water and low salinity. This correlates with *in vitro* research showing *B.ps* counts dropping
70 rapidly in salt concentrations above 2.5% (9) or seawater (16). Soft water can be corrosive and
71 compromise bore piping, potentially facilitating *B.ps* introduction into bores and creating a rough
72 inner bore surface promoting biofilms (1).

73 Coliform counts were significantly higher in *B.ps* positive bores suggesting the presence of
74 nutrients for microbial growth. There was also a significant association between increased
75 turbidity and *B.ps*. Higher turbidity likely reflects failure of the bore casing in the subterranean
76 environment or backflow of surface water into the bore. Contamination of the bore with soil may
77 well explain the origins of *B.ps* and other microbes in these bores, but this requires formal study.
78 More organic matter in these bores would be favourable to the saprophytic *B.ps* and
79 decomposition of organic matter would contribute to acidification of the water.

80 The occurrence of *B.ps* was strongly associated with high iron levels. This finding supports
81 previous research showing enhanced *B.ps* growth in iron-rich media (19) and red-coloured soil
82 (indicating oxidized iron) (11). Clinical conditions causing iron overload such as thalassemia are
83 associated with increased melioidosis rates (3) and *B.ps* is able to produce siderophores under
84 limited iron supply (19). It is of interest that acidic pH associated with *B.ps* positive bores
85 increases bio-availability of iron by reduction of precipitated Fe^{3+} to soluble Fe^{2+} , especially
86 under more anaerobic conditions such as if water is pumped up from aquifers (12).

87 A comparison of dry and wet season data showed that water parameters in *B.ps* positive bores
88 were even more favourable for *B.ps* in the wet season. Heavy rainfall might explain reduced
89 salinity in the wet season, with higher iron levels due to water influx from shallow aquifers in an
90 iron-rich lateritic layer which are only active in the wet season.

91 Clustered multivariable analysis showed that the most significant predictors for presence of *B.ps*
92 in water were high iron levels and the interaction of a low pH with low salinity (Table 1).

93

94 **Conclusion.** We have found a close association between *B.ps* presence in bore water and water
95 parameters such as low pH, low salinity and high iron levels. This indicates that *B.ps* occurrence
96 in bores is not only the result of an initial contamination event but also depends on water
97 conditions favourable for *B.ps*. The strong association of *B.ps* with an abiotic fingerprint aids in
98 identification of water bores at high risk of *B.ps* contamination. Future studies will examine
99 whether changing levels of abiotic parameters such as through pH-correction filters create an
100 environment unfavourable for *B.ps* growth and thus, could be used as a preventive measure
101 against *B.ps* persistence in unchlorinated water supplies.

102

103 **Acknowledgments** We would like to thank the Darwin rural community for access to their water
104 bores. We are grateful to Leisha Richardson for assistance with MLV-4 work and the staff of
105 NRETA, particularly to Kevin Boland for advice on hydrogeology and water bores. We thank
106 Joseph McDonnell for helpful comments regarding statistical analysis. This work was supported
107 in part by an Australian National Health and Medical Research Council Project Grant 383504 (to
108 BJC and MM) and the Swiss National Science Foundation (to MK).

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References

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- 111 1. **Australian Government National Health and Medical Research Council.** 2006.
112 Australian Drinking Water Guidelines 6.
113 http://www.nhmrc.gov.au/_files_nhmrc/file/publications/synopses/adwg_11_06.pdf:accessed December 27th 2009.
114
- 115 2. **Chen Y.S., S.C.Chen, C.M.Kao, and Y.L.Chen.** 2003. Effects of soil pH, temperature
116 and water content on the growth of *Burkholderia pseudomallei*. *Folia Microbiol (Praha)*.
117 **48**:253-256.
- 118 3. **Cheng A.C., B.J.Currie.** 2005. Melioidosis: epidemiology, pathophysiology, and
119 management. *Clin Microbiol Rev.* **18**:383-416.
- 120 4. **Currie B.J., D.A.Fisher, D.M.Howard, J.N.Burrow, S.Selvanayagam, P.L.Snelling,**
121 **N.M.Anstey, and M.J.Mayo.** 2000. The epidemiology of melioidosis in Australia and
122 Papua New Guinea. *Acta Trop.* **74**:121-127.
- 123 5. **Currie B.J., M.Mayo, N.M.Anstey, P.Donohoe, A.Haase, and D.J.Kemp.** 2001. A
124 cluster of melioidosis cases from an endemic region is clonal and is linked to the water
125 supply using molecular typing of *Burkholderia pseudomallei* isolates. *Am J Trop Med Hyg.*
126 **65**:177-179.

- 127 6. **Haig T, Townsend S.** 2003. An understanding of the groundwater and surface water
128 hydrology of the Darwin Harbour Plan of Management area. Proceedings of the Darwin
129 Harbour Public Presentations.
- 130 7. **Inglis T.J., N.F.Foster, D.Gal, K.Powell, M.Mayo, R.Norton, and B.J.Currie.** 2004.
131 Preliminary report on the northern Australian melioidosis environmental surveillance
132 project. *Epidemiol Infect.* **132**:813-820.
- 133 8. **Inglis T.J.J., S.C.Garrow, M.Henderson, A.Clair, J.Sampson, L.O'Reilly, and**
134 **B.Cameron.** 2000. *Burkholderia pseudomallei* Traced to Water Treatment Plant in
135 Australia. *Emerging Infectious Diseases.* **6**:56-59.
- 136 9. **Inglis T.J.J., J.L.Sagripanti.** 2006. Environmental Factors That Affect the Survival and
137 Persistence of *Burkholderia pseudomallei*? *Appl Envir Microbiol.* **72**:6865-6875.
- 138 10. **Kaestli M., M.Mayo, G.Harrington, L.Ward, F.Watt, J.Hill, A.C.Cheng, and**
139 **B.J.Currie.** 2009. Landscape Changes Influence the Occurrence of the Melioidosis
140 Bacterium *Burkholderia pseudomallei* in Soil in Northern Australia. *PLOS Neglected*
141 *Tropical Diseases.* **3(1): e364. doi:10.1371/journal.pntd.0000364:**
- 142 11. **Kaestli M., M.Mayo, G.Harrington, F.Watt, J.Hill, D.Gal, and B.J.Currie.** 2007.
143 Sensitive and Specific Molecular Detection of *Burkholderia pseudomallei*, the Causative
144 Agent of Melioidosis, in the Soil of Tropical Northern Australia. *Appl Envir Microbiol.*
145 **73**:6891-6897.
- 146 12. **Manahan S.E.** 2010. *Environmental Chemistry.* **7th Edition:**Lewis Publishers London.

- 147 13. **Northern Territory Government Department of Natural Resources E. a. t. A.** 2008.
148 Groundwater Supply Prospects and Hydrogeology of the Litchfield Shire (Map).
- 149 14. **Novak R.T., M.B.Glass, J.E.Gee, D.Gal, M.J.Mayo, B.J.Currie, and P.P.Wilkins.**
150 2006. Development and evaluation of a real-time PCR assay targeting the type III secretion
151 system of *Burkholderia pseudomallei*. *J Clin Microbiol.* **44**:85-90.
- 152 15. **Palasatien S., R.Lertsirivorakul, P.Royros, S.Wongratanacheewin, and**
153 **R.W.Serm Swan.** 2008. Soil physicochemical properties related to the presence of
154 *Burkholderia pseudomallei*. *Trans R Soc Trop Med Hyg.* **102 Suppl 1**:S5-S9.
- 155 16. **Robertson J., A.Levy, J.L.Sagripanti, and T.J.Inglis.** 2010. The survival of *Burkholderia*
156 *pseudomallei* in liquid media. *Am J Trop Med Hyg.* **82**:88-94.
- 157 17. **Strauss J.M., M.G.Groves, M.Mariappan, and D.W.Ellison.** 1969. Melioidosis in
158 Malaysia. II. Distribution of *Pseudomonas pseudomallei* in soil and surface water. *Am J*
159 *Trop Med Hyg.* **18**:698-702.
- 160 18. **Tong S., S.Yang, Z.Lu, and W.He.** 1996. Laboratory investigation of ecological factors
161 influencing the environmental presence of *Burkholderia pseudomallei*. *Microbiol Immunol.*
162 **40**:451-453.
- 163 19. **Yang H.M., W.Chaowagul, and P.A.Sokol.** 1991. Siderophore production by
164 *Pseudomonas pseudomallei*. *Infect Immun.* **59**:776-780.
165

166 **TABLE 1** Summary of Water Parameters

167

Variable	Dry Season		Wet Season		Multivariable Analysis
	Median (95% CI)	Mann-Whitney	Median (95% CI)	Mann-Whitney	OR (95% CI) P value
** Total Iron (mg/L)					
<i>B.ps</i> positive	2 (1-3)	P<0.001	4 (2-5)	P=0.002	1.34 (1.12-2.62) P=0.002
<i>B.ps</i> negative	1 (1-1)		1 (1-1)		
** pH					
<i>B.ps</i> positive	6.8 (6.5-7.1)	P<0.001	6.3 (6.1-6.6)	P<0.001	Interaction of pH and Salinity
<i>B.ps</i> negative	7.3 (7.2-7.4)		7.4 (7.3-7.6)		
* Salinity (mS/cm)					<0.01 (<0.01-0.36) P=0.027
<i>B.ps</i> positive	0.07 (0.02-0.25)	P=0.037	0.02 (0.01-0.05)	P<0.001	
<i>B.ps</i> negative	0.25 (0.17-0.27)		0.25 (0.09-0.28)		
* Hardness (mg/L)					
<i>B.ps</i> positive	100 (40-170)	P=0.066	25 (10-80)	P<0.001	
<i>B.ps</i> negative	180 (150-200)		170 (80-180)		
Phosphate (mg/L)					
<i>B.ps</i> positive	2 (1-3)	P=0.114	6 (4-6)	P=0.069	
<i>B.ps</i> negative	3 (2-3)		3 (3-6)		
** Turbidity (NTU)					
<i>B.ps</i> positive	14.0 (6.0-22.0)	P<0.001	15.0 (10.0-26.0)	P=0.001	
<i>B.ps</i> negative	3.1 (2.6-4.0)		6.4 (5.0-8.0)		
* Coliforms (CFU/mL)					
<i>B.ps</i> positive	100 (20-220)	P=0.033	120 (21-220)	P=0.004	
<i>B.ps</i> negative	12 (5-25)		9 (2-24)		
Effective Bore Capping	<i>B.ps</i> positive	Fisher's Exact	<i>B.ps</i> positive	Fisher's Exact	
Yes (38 bores)	21 %	P=0.205	29 %	P=0.438	0.38 (0.14-1.06) P=0.067
No (9 bores)	44 %		44 %		

168

169

170 **TABLE 1:** Summary statistics of the occurrence of *B.ps* in water bores and bore parameters.
171 Using Stata/IC 10.0 (USA), 95% CI are percentile bootstrap estimates, odds ratios for *B.ps*
172 positive bores were calculated using multivariable logistic regression clustered by bore and
173 allowing standard errors for intra-group correlation and including season. The model was
174 specified correctly as tested by a linktest. * indicates statistically significant result with $P < 0.01$ in
175 either dry or wet season and ** a significant result in dry and wet season. Nitrate levels are not
176 reported as most were below test detection levels of 10 mg/L.

177 **FIGURE LEGEND**

178

179 **FIGURE 1:** Box-and-whisker plots of water parameters in B.ps negative (“neg”) and positive
180 (“pos”) bores of the dry and wet season. The box spans the interquartile range of the data and the
181 median is marked with a vertical line. * indicates a significant difference between B.ps negative
182 and positive bores with $P < 0.01$ (Mann-Whitney U test). Unit of salinity (conductivity) is
183 mS/cm (mS/cm x 640 = ppm total salts).

184

Box plots of water parameters of *B. pseudomallei* positive and negative bores

